RANDOMIZED PROSPECTIVE DOUBLE-BLIND PLACEBO-CONTROLLED STUDY OF DEXTROSE PROLOOTHERAPY FOR KNEE OSTEOARTHRITIS WITH OR WITHOUT ACL LAXITY

Kenneth D. Reeves, MD, and Khatab Hassanein, PhD

Kenneth D. Reeves is the section chief for Physical Medicine and Rehabilitation at Bethany Medical Center, Kansas City, Kan. Khatab Hassanein is professor and chairman for the Department of Biometry at the University of Kansas Medical Center, Kansas City, Kan.

Context • Use of prolotherapy (injection of growth factors or growth factor stimulators).
Objective • Determine the effects of dextrose prolotherapy on knee osteoarthritis with or without anterior cruciate ligament (ACL) laxity.
Design • Prospective randomized double-blind placebo-controlled trial.
Setting • Outpatient physical medicine clinic.
Patients or other participants • Six months or more of pain along with either grade 2 or more joint narrowing or grade 2 or more osteophytic change in any knee compartment. A total of 38 knees were completely void of cartilage radiographically in at least 1 compartment.
Intervention • Three bimonthly injections of 9 cc of either 10% dextrose and .075% lidocaine in bacteriostatic water (active solution) versus an identical control solution absent 10% dextrose. The dextrose-treated joints then received 3 further bimonthly injections of 10% dextrose in open-label fashion.
Main Outcome Measures • Visual analogue scale for pain and swelling, frequency of leg buckling, goniometrically measured flexion, radiographic measures of joint narrowing and osteophytosis, and KT1000-measured anterior displacement difference (ADD).
Results • All knees: Hotelling multivariate analysis of paired observations between 0 and 6 months for pain, swelling, buckling episodes, and knee flexion range revealed significantly more benefit from the dextrose injection (P = .015). By 12 months (6 injections) the dextrose-treated knees improved in pain (44% decrease), swelling complaints (63% decrease), knee buckling frequency (85% decrease), and in flexion range (14 degree increase). Analysis of blinded radiographic readings of 0- and 12-month films revealed stability of all radiographic variables except for 2 variables which improved with statistical significance. (Lateral patellofemoral cartilage thickness [P = .019] and distal femur width in mm [P = .021]. Knees with ACL laxity: 6-month (3 injection) data revealed no significant improvement. However, Hotelling multivariate analysis of paired values at 0 and 12 months for pain, swelling, joint flexion, and joint laxity in the dextrose-treated knees, revealed a statistically significant improvement (P = .021). Individual paired t tests indicated that blinded measurement of goniometric knee flexion range improved by 12.8 degrees (P = .005), and ADD improved by 57% (P = .025). Eight out of 13 dextrose-treated knees with ACL laxity were no longer lax at the conclusion of 1 year.
Conclusion • Prolotherapy injection with 10% dextrose resulted in clinically and statistically significant improvements in knee osteoarthritis. Preliminary blinded radiographic readings (1-year films, with 3-year total follow-up period planned) demonstrated improvement in several measures of osteoarthritic severity. ACL laxity, when present in these osteoarthritic patients, improved. (Altern Ther Health Med. 2000;6(1):68-80)

INTRODUCTION

Prolotherapy (injection of growth factors or growth factor stimulators) raises growth factor levels or increases growth factor effectiveness to promote tissue repair or growth. The most common solutions used for prolotherapy create a brief inflammatory response. Temporary cellular stress causes a release of cytokines and increased growth factor activity with migration of macrophages (white blood cells), and then multiplication of repair cells specific to the tissue. Unlike repair after an injury, disruption of architecture of tissue from injury does not occur, and new cells and matrix can be deposited in an organized fashion, with maturation of new tissue for 6 to 8 weeks. Two double-blind studies have been performed on prolotherapy in low back pain using inflammatory solutions. These studies both showed significant benefit from proliferant injection, but because the solutions were inflammatory there was some potential for impairment of double-blind protocol. The purpose of this investigation was to evaluate effectiveness of prolotherapy without using any inflammatory mechanism so that neither patient, research coordinator
nor primary investigator would have any way to determine patient group. Our specific plan was to study the effect of a non-inflammatory (10%) concentration of dextrose (D-glucose in water) on knee osteoarthritis patients via objective measures of knee cartilage, knee osteophytic status, and knee goniometric range, as well as by subjective measures of knee pain, knee swelling, and knee buckling.

Some patients in the study had anterior cruciate ligament (ACL) laxity, which is known to initiate and worsen knee osteoarthritis. A second purpose for this study was to observe the effect of proliferant injection on laxity of the ACL, as measured by an objective and reproducible measure (an electroarthrometer).

Elevation of extracellular glucose to as little as .5% (normal extracellular and cellular glucose is .1%) has been shown to raise levels of multiple polypeptide growth factors in a variety of human cells. Exposure of several human cells to a hypertonic environment will also promptly result in a rise in DNA levels for growth factors within seconds to minutes. Therefore, hypertonic dextrose solution has 2 mechanisms by which to increase levels of growth factors, potentially improving the status of critical cells in the joint such as chondrocytes (cartilage producing cells), osteocytes (bone producing cells), and fibroblasts (ten-}

tomological criteria for knee osteoarthritis, they were assigned serially to group 1 or 2 using a random number table by 1 of 2 data base coordinators always in the office. This group assignment was kept in a database blinded to the chief investigator and research coordinator.

The research coordinator obtained an estimate of arthritis medications taken and then demonstrated the use of a 100-mm visual analogue scale (VAS) and gave 3 examples of its use. The patients then self-scored their pain levels of knee pain at rest, knee pain walking on level surfaces, knee pain with stair use, and subjective swelling, and estimated the number of knee buckling episodes over the previous 2 months. Following this, the research coordinator obtained goniometric readings of joint flexion by the method described in a standard text. Patients who were taking any medication or oral supplement for osteoarthritis other than calcium, multivitamins, NSAIDS, acetaminophen, or occasional narcotic, were asked to discontinue them. The most common oral supplement discontinued was glucosamine/chondroitin sulfate.

Blood was obtained for sedimentation rate, rheumatoid factor, uric acid, and antinuclear antibody. Significant laboratory abnormalities led to referral to primary physician or rheumatologist for determination of the presence or absence of inflammatory arthritis. No patients required exclusion due to the laboratory battery after the initial phone screening.

Dextrose prolotherapy solutions for maximum safety have typically included bacteriostatic water, a small concentration of lidocaine, and dextrose. Because of the desire to maximize safety and comfort in this study and simulate typical prolotherapy solutions, the control was the usual bacteriostatic water with a very small amount of lidocaine, and the active solution was identical except for the inclusion of 10% dextrose.

At 0, 2, and 4 months solution was drawn up blinded to both chief investigator and research coordinator. Using a 27-gauge needle via an interferomedial approach, tibiofemoral injection was conducted with 9 cc of either 611.4 mOsm (10% dextrose and .075% lidocaine in bacteriostatic water) or 105.4 mOsm (.075% lidocaine in bacteriostatic water) solution. Bacteriostatic water consisted of .9% benzyl alcohol. The small dose of lidocaine was included for postinjection comfort. The solutions were identical in color and viscosity. Dextrose at 10% concentration is very slightly sticky if allowed to dry on the skin but Hibiclens was used for glove and skin prep which masked any potential of noting any slight stickiness of solution.

Treatment continued beyond 6 months in the dextrose group with additional injections at 6, 8, and 10 months. Subjective variables, goniometric flexion, 2-view radiographs and KT-1000 ADD measurements were repeated at 1 year. Skier’s (standing) views of the knee were used to determine tibiofemoral compartment status. Angle of knee flexion on skier’s views, angle of radiograph beam to the knee, camera to film distance, power and duration of radiograph beam, and radiograph technician were identical at 0 and 12 months. Magnification on standing films was prevented by ensuring contact of patella with film plate. Skyline views of the patella
were used to determine patellofemoral compartment status, with similar measurements, including camera to knee and knee to film distance, to ensure an identical radiograph method.

Radiographs were read in double-blind fashion in the following way. The study coordinator obscured patients’ names and labeled the film with a random patient number. The film date was obscured and a random number table was used to assign a number to the 0- and 12-month films. The 0- and 12-month films were then separated in different packets so that reading 1 film would not influence reading of the next. Osteophytic grade was measured in 6 compartments using a standard atlas with approximately 90% intra-reader agreement.12 The compartments included medial femoral, medial tibial, lateral femoral, lateral tibial, medial patellofemoral, and lateral patellofemoral. Cartilage thickness was determined in 4 compartments in millimeters: medial tibiofemoral, lateral tibiofemoral, medial patellofemoral, and lateral patellofemoral. General hypertrophic change was evaluated as a width measurement in millimeters: distal femur width proximal to the intercondylar notch, distal femur width distal to the intercondylar notch, and proximal tibial width. Width measurements were made parallel with the film bottom edge through the area of largest width, including any osteophytes present. The x-rays were read by the chief investigator. A database coordinator loaded results onto the database.

Human subject research approval and monitoring was by the Institutional Review Committee of Bethany Medical Center in Kansas City, Kans. Procedures followed were in accordance with ethical standards outlined in the Helsinki Declaration Revision of 1983. The statistical analysis software was SPSS (Statistical Program for Social Science) version 7.5.3.

RESULTS

Blinding method problems were not identified. No treatment complications were noted. Seventy-seven patients had 1 or more knees that met study criteria for symptomatic osteoarthritis (OA). Nine patients dropped out over 12 months of followup, 4 due to lack of efficacy (3 in control group and 1 in active group), and 5 for unrelated medical reasons. This left 111 knees in 68 patients with OA.

At study onset 31 patients met arthrometric criteria for ACL laxity. Two dropped out over 12 months due to lack of efficacy and 4 for unrelated medical issues, leaving 25 for analysis. Independent sample t tests were conducted to compare the active and control groups of OA knees. No significant differences were noted between groups for age, weight, pain levels, range of motion, buckling episodes, or radiographic findings. The same result was noted when t tests were conducted to compare active and control groups of knees with ACL laxity. The average knee OA patient in this study was 63 years of age and weighed 195 lb. Males comprised 58% of the study population.

Complications and Safety Issues

Discomfort after injection did not appear to vary between groups, typically lasting a few minutes to several days. Despite use of an allergy-size needle (27 gauge) and a single-insertion technique, some patients had pain with distension of the joint capsule even with this minimal volume (9 cc). The 9 cc volume for injection may be a bit excessive in that some patients were inhibited in flexion for several days. One person had a flare postinjection that appeared substantial, requiring interarticular steroid and then referral to an orthopedic surgeon. When blinding was broken she was found to have received control solution.

No allergic reactions or infections were noted.

Six-Month (Double-Blind Phase) Data Comparing Active and Control Solution for all Osteoarthritic Knees

Figure 1 presents a bar graph depicting improvements in pain (average of improvement in pain at rest, pain with walking, and pain with stair use) and swelling for the active and control groups at 6 months (after 3 injections of 9 cc of solution). Both active and hypotonic control solution administration resulted in considerable gains in VAS scores for pain.

Figure 2 shows improvement in knee flexion for both groups at 6 months. Both active and hypotonic control injections resulted in an improvement in goniometric knee flexion measures.

Hotelling multivariate analysis of paired observations between 0 and 6 months for active and control solution including all nonradiographic variables (pain at rest, pain with walking, pain with stair use, swelling, buckling episodes, and flexion range) demonstrated a statistically superior effect of active solution (P=.015). The results of individual paired t tests from 0 to 6

![FIGURE 1 Percentage improvement in pain and swelling from study onset to 6 months for active and control solutions.](image-url)
months for each of the variables are shown in Table 1. Although the active solution was superior statistically, highly significant improvement from 0 to 6 months was seen in pain with walking, pain with stair use, and flexion range of motion in both active and control groups.

The NSAID follow-up question was limited in its ability to determine a degree of change in level of intake, and no significant change between groups was noted. However, neither group had an increase in NSAID intake, which could explain improvement in pain levels or other variables.

One-Year Data (Nonradiographic) for Active Solution for Osteoarthritic Knees

Figure 3 shows percentage improvements in pain and swelling complaints and knee buckling in the dextrose group between 0 and 12 months (with 3 further bimonthly open label injections of dextrose). Pain improved by 40%, swelling by 63%, buckling episodes by 85%, and flexion by 14 degrees as compared with study entry.

Radiographic Data at 1 Year for Active Solution for Osteoarthritic Knees (Table 2)

Thirteen radiographic readings for each knee are shown in Table 2. These variables included medial femoral osteophyte grade (MFOG), medial tibial osteophyte grade (MTOG), lateral femoral osteophyte grade (LFOG), lateral tibial osteophyte grade (LTOG), medial patellofemoral osteophyte grade (MPOG), lateral patellofemoral osteophyte grade (LPOG), medial tibiofemoral cartilage thickness (MTFT), lateral tibiofemoral cartilage thickness (LTFT), medial patellofemoral cartilage thickness (MPFT), lateral patellofemoral cartilage thickness (LPFT), distal femur width proximal to the intercondylar notch (DFWP), distal femur width distal to the intercondylar notch (DFWD), and proximal tibial width (PTW).

Hotelling multivariate analysis of paired observations between 0 and 12 months for the dextrose-treated knees including all 13 radiographic variables revealed a statistically significant change ($P = .028$). Individual paired $t$ tests showed the means for radiographic variables were all stable except for an improvement (increase) in lateral patellofemoral cartilage thickness ($P = .019$) and an improvement (decrease) in distal femur width including osteophytes ($P = .021$).

Data for Knees with ACL Laxity

The 6-month data showed no statistically significant differences between active and control solutions, nor significant changes in ACL laxity measurement. However, the dextrose-treated knees were given 3 additional injections of dextrose and data were collected at 1-year follow-up. Hotelling multivariate analysis of paired observations of the dextrose-treated knees comparing 0 and 12 months for VAS rest pain, VAS walking pain, VAS stair use pain, VAS swelling complaint, flexion range of motion, and KT1000 side-to-side difference showed statistically significant improvement over time ($P = .021$). The results of individual paired $t$ tests from 0 to 12 months are shown in Table 3. Blinded goniometric range measurement improved by 12.8 degrees with a $P$ value of .005 and KT1000 ADD improved by 57% with a $P$ value of .025. Figure 4 is a bar graph showing the
distribution frequency of laxity values (ADD) for dextrose-treated patients at time 0 and 12 months. Note that 8 of the 13 improved to the point that ADD was less than 2, such that they would no longer be considered lax, and this in a group of patients with one or more complete ACL ruptures.

**DISCUSSION**

**Balance of Growth and Disrepair Factors in Bony Cortex, Cartilage, and Synovial Fluid**

The balance of disrepair and repair in both bone and cartilage merit examination since degenerative changes in bone occur simultaneously with those in cartilage.\textsuperscript{21,22} Chief repair factors found in osteoarthritic subchondral bone or cartilage include insulin-like growth factor (IGF), transforming growth factor beta (TGF-\(\beta\)), epidermal growth factor (EGF), basic fibroblast growth factor (bFGF), and platelet derived growth factor (PDGF).\textsuperscript{23-27} Chief disrepair factors (factors that block growth factor effects or break down tissue or building blocks for tissue) for the bony surface or cartilage include interleukin-1 (IL-1) and tumor necrosis factor (TNF), which lead to a rise as much as 110-fold in metalloproteinases such as collagenase (which breaks down cartilage) in fibrillated cartilage and a rise as much as 24-fold in binding proteins (proteins that bind growth factors to keep them from functioning) in synovial fluid.\textsuperscript{23-27}

### TABLE 1  Means, standard deviations (SD), and individual paired t tests for change in nonradiographic variables from 0 to 6 months in all osteoarthritic knees for active and control solution

<table>
<thead>
<tr>
<th>Group</th>
<th>Mean (SD) 0 months</th>
<th>Mean (SD) 6 months</th>
<th>Mean diff 0-6 months</th>
<th>Standard error of mean diff</th>
<th>95% CI for the mean difference</th>
<th>Significance between means at 0 and 6 months</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pain at rest</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Active</td>
<td>2.15 (2.24)</td>
<td>1.61 (1.71)</td>
<td>-.54</td>
<td>.24</td>
<td>-1.02 to -.06</td>
<td>.029</td>
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<tr>
<td>Control</td>
<td>2.73 (2.02)</td>
<td>1.69 (1.73)</td>
<td>-1.04</td>
<td>.25</td>
<td>-1.54 to -.54</td>
<td>.00005</td>
</tr>
<tr>
<td>Pain with walking</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Active</td>
<td>3.94 (2.82)</td>
<td>2.56 (1.97)</td>
<td>-1.39</td>
<td>.31</td>
<td>-2.01 to -.77</td>
<td>.00002</td>
</tr>
<tr>
<td>Control</td>
<td>3.83 (2.20)</td>
<td>2.85 (2.20)</td>
<td>-.98</td>
<td>.32</td>
<td>-1.62 to -.34</td>
<td>.003</td>
</tr>
<tr>
<td>Pain with stair use</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Active</td>
<td>5.33 (2.80)</td>
<td>3.96 (2.68)</td>
<td>-1.37</td>
<td>.32</td>
<td>-2.01 to -.73</td>
<td>.00004</td>
</tr>
<tr>
<td>Control</td>
<td>5.83 (2.60)</td>
<td>4.60 (2.91)</td>
<td>-1.23</td>
<td>.32</td>
<td>-1.87 to -.59</td>
<td>.0002</td>
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<tr>
<td>Swelling</td>
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<tr>
<td>Active</td>
<td>2.44 (2.53)</td>
<td>1.35 (1.87)</td>
<td>-1.09</td>
<td>.25</td>
<td>-1.59 to -.59</td>
<td>.00003</td>
</tr>
<tr>
<td>Control</td>
<td>3.12 (2.99)</td>
<td>2.52 (2.80)</td>
<td>-.60</td>
<td>.26</td>
<td>-1.12 to -.08</td>
<td>.022</td>
</tr>
<tr>
<td>Buckling episodes per 2 months</td>
<td>7.78 (34.14)</td>
<td>2.54 (11.44)</td>
<td>-5.24</td>
<td>2.23</td>
<td>-9.70 to -.78</td>
<td>.020</td>
</tr>
<tr>
<td>Control</td>
<td>1.00 (2.60)</td>
<td>.21 (.64)</td>
<td>-.79</td>
<td>2.27</td>
<td>-5.33 to +3.75</td>
<td>.729</td>
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<tr>
<td>Flexion range</td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Active</td>
<td>112.35 (19.54)</td>
<td>125.59 (8.63)</td>
<td>-13.24</td>
<td>2.13</td>
<td>+8.94 to +17.54</td>
<td>.00000001</td>
</tr>
<tr>
<td>Control</td>
<td>117.75 (11.32)</td>
<td>123.44 (7.48)</td>
<td>-7.69</td>
<td>2.19</td>
<td>+3.31 to +12.07</td>
<td>.001</td>
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</table>
Table 2: Means, standard deviations (SD), and individual paired *t* tests for change in radiographic variables from 0 to 12 months in osteoarthritic knees treated with active solution

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mean (SD) 0 months</th>
<th>Mean (SD) 6 months</th>
<th>Mean diff 0 - 12 months</th>
<th>Standard error of mean diff</th>
<th>95% CI for the mean difference</th>
<th>Significance between means at 0 and 6 months</th>
<th>Direction of change</th>
</tr>
</thead>
<tbody>
<tr>
<td>MFOG</td>
<td>1.55 (1.07)</td>
<td>1.49 (1.02)</td>
<td>-.06</td>
<td>.11</td>
<td>-28 to +.16</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>MTOG</td>
<td>1.56 (.92)</td>
<td>1.53 (1.05)</td>
<td>-.03</td>
<td>.10</td>
<td>-23 to +1.7</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>LFOG</td>
<td>1.65 (.91)</td>
<td>1.76 (.84)</td>
<td>+.11</td>
<td>.13</td>
<td>-15 to +.37</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>LTOG</td>
<td>1.22 (.95)</td>
<td>1.33 (1.00)</td>
<td>+.11</td>
<td>.13</td>
<td>-15 to +.37</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>MPOG</td>
<td>1.24 (.82)</td>
<td>1.25 (.82)</td>
<td>-.01</td>
<td>.12</td>
<td>-25 to +.23</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>LPOG</td>
<td>1.42 (.66)</td>
<td>1.40 (.66)</td>
<td>-.02</td>
<td>.08</td>
<td>-18 to +.14</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>MTFT</td>
<td>2.09 (2.18)</td>
<td>1.94 (2.14)</td>
<td>-.15</td>
<td>.13</td>
<td>-41 to +.11</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>LTFT</td>
<td>5.54 (2.06)</td>
<td>5.58 (2.32)</td>
<td>+.04</td>
<td>.17</td>
<td>-.30 to +.38</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>MPFT</td>
<td>4.51 (1.63)</td>
<td>4.59 (1.41)</td>
<td>+.08</td>
<td>.19</td>
<td>-.30 to +.46</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>LPFT</td>
<td>4.20 (1.54)</td>
<td>4.59 (1.34)</td>
<td>+.39</td>
<td>.16</td>
<td>+.07 to +.71</td>
<td>.019 Improved</td>
<td>Improved</td>
</tr>
<tr>
<td>DFWP</td>
<td>93.58 (7.23)</td>
<td>92.96 (7.07)</td>
<td>-.62</td>
<td>.26</td>
<td>-1.14 to -.10</td>
<td>.021 Improved</td>
<td>Improved</td>
</tr>
<tr>
<td>DFWD</td>
<td>90.18 (8.36)</td>
<td>90.60 (8.14)</td>
<td>+.42</td>
<td>.34</td>
<td>-26 to +1.10</td>
<td>NS</td>
<td>Stable</td>
</tr>
<tr>
<td>PTW</td>
<td>89.18 (7.96)</td>
<td>88.53 (8.08)</td>
<td>-.65</td>
<td>.39</td>
<td>-1.43 to +.13</td>
<td>NS</td>
<td>Stable</td>
</tr>
</tbody>
</table>

Proliferation of Human Chondrocytes by Growth Factors in Culture and Chondrogenesis of Animal Cartilage by Injection of Growth Factors

Bujia and Dunham demonstrated that culturing human chondrocytes (nasal septum chondrocytes) in fluid containing TGF-β, IGF-1, or bFGF resulted in proliferation. Injection of animal knees with a single injection of TGF-β, bone metabolic protein-2 (BMP-2), bFGF, or hepatocyte growth factor (HGF) has led to chondrogenesis, enlargement of articular cartilage, and repair of full thickness joint cartilage defects. Implantation of gel or a collagen sponge saturated with growth factor or placement of a surgically placed small pump that delivers growth factors have led to repair of full thickness cartilage lesions in animal models. However, demonstration of weeks of proteoglycan synthesis after a single injection of TGF-β indicates that continuous exposure to growth factor may not be required for a prolonged growth factor effect. In established OA high levels of binding proteins or metalloproteinases may block the effect of a single growth factor.
injection, and there may be a need in humans to combine growth factors with agents that neutralize disrepair factors for optimum effectiveness in established osteoarthritis.\textsuperscript{35}

### Table 3

<table>
<thead>
<tr>
<th>Mean (SD) 0 months</th>
<th>Mean (SD) 12 months</th>
<th>Mean diff 0-12 months</th>
<th>Standard error of mean diff</th>
<th>95% CI for the mean difference</th>
<th>Significance between means at 0 and 12 months</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pain at rest</td>
<td>2.31 (2.56)</td>
<td>1.38 (2.06)</td>
<td>-0.93 (0.49)</td>
<td>-1.91 to +0.06</td>
<td>0.082</td>
</tr>
<tr>
<td>Pain with walking</td>
<td>3.77 (2.77)</td>
<td>2.31 (2.72)</td>
<td>-1.46 (0.46)</td>
<td>-2.38 to -0.92</td>
<td>0.008</td>
</tr>
<tr>
<td>Pain with stair use</td>
<td>5.54 (3.31)</td>
<td>4.15 (3.29)</td>
<td>-1.39 (0.47)</td>
<td>-2.33 to -0.45</td>
<td>0.013</td>
</tr>
<tr>
<td>Swelling</td>
<td>2.77 (2.71)</td>
<td>1.54 (2.40)</td>
<td>-1.23 (0.66)</td>
<td>-2.55 to +0.09</td>
<td>0.008</td>
</tr>
<tr>
<td>Flexion range</td>
<td>112.69 (16.93)</td>
<td>125.46 (6.89)</td>
<td>+12.77 (3.77)</td>
<td>+5.23 to +20.31</td>
<td>0.005</td>
</tr>
<tr>
<td>KT1000 side to side diff</td>
<td>3.08 (1.32)</td>
<td>1.23 (2.24)</td>
<td>-1.85 (0.72)</td>
<td>-3.29 to -0.41</td>
<td>0.005</td>
</tr>
</tbody>
</table>

### Effect on Human Cells of Exposure to Elevated Glucose

Elevation of extracellular glucose to as little as .5% has been shown to raise levels of IGF-1 in human mesangial (glomerular) cells,\textsuperscript{6} IGF-2 in human mesangial cells,\textsuperscript{7} TGF-\(\beta\)1 in human mononuclear cells\textsuperscript{6} and human mesangial cells,\textsuperscript{6,7} PDGF-B (platelet derived growth factor beta) in human mesangial cells\textsuperscript{6} and human capillary endothelial cells,\textsuperscript{37} bFGF in human gingival fibroblasts,\textsuperscript{6} and connective tissue growth factor (CTGF) in human mesangial cells.\textsuperscript{3} In addition, glucose in blood mononuclear cells has been found to suppress potential disrepair factors (interleukins such as IL-2, IL-6, and IL-10).\textsuperscript{3} Cellular response to elevated extracellular glucose is swift. DNA levels for growth factor production rise within minutes to hours of cellular exposure to elevated glucose concentrations.\textsuperscript{3} As many as 15 different genes are induced with exposure to elevated glucose concentration.\textsuperscript{3}

### Effect on Human Cells of Exposure to Osmolar Changes

Exposure of a cell to an osmolarity change as little as 50 mOsm has also been found to activate enzymes (phosphate donors, also termed kinases) in the cell similar to the growth factors mentioned above.\textsuperscript{4,5,38-40} The mechanism appears to be via a
change in cell size, leading to kinase production via natural cellular responses to stress. Although the kinases produced by osmolar change are not the same as with glucose elevation, proliferation response to a change in osmolarity has been demonstrated and at least 1 kinase produced is clearly a growth factor related to proliferation (PDGF).

**Potential Therapeutic Benefit of Bacteriostatic Water Solution or Anesthetic**

The osmolarity of the bacteriostatic water solution used for the control injection was 105 compared to 611 for the active group. Information about the potential efficacy of hypotonic solution came out in the literature after our study began. This raised the question of whether the hypotonic control solution in this study was more than a placebo treatment. Review of placebo responses in recent double-blind studies of knee osteoarthritis revealed a range of pain reduction from 9% to 30%. Review of studies on knee OA in which knee flexion measurements were obtained before and after treatment yielded few studies. A search over the last 30 years indicated a range of improvement in knee flexion in placebo groups from a -4.6 degree loss to a 1 degree gain. The control group in this study improved by 28% in pain and 8 degrees in flexion range, suggesting more than a placebo effect. Since ligaments in different locations in animals respond to different growth factors there may be dissimilar findings for different joints. Thus it is of interest that a concurrent finger OA study did show similar benefit with dextrose solution but the control solution did not show an appreciable benefit. It is possible that there was some therapeutic effect from inclusion of anesthetic in the solution as well, and if so this may explain some benefit in both groups. However, the concentration of lidocaine at 0.075% was quite low and was identical in both treatment solutions.

**Magnitude of Clinical Benefit from Dextrose Solution Use Compared to Active Groups in Other Recent Studies**

Pain improvement in the active treatment group by 40% through 1 year after 6 injections of 9 cc of simple dextrose solution approximated that of the active treatment group in recent studies on avocado soybean unsaponifiables, chondroitin sulfate, glucosamine, and NSAIDs. Range of motion improvement in flexion in the dextrose-treated knees (14 degrees) exceeded the range of flexion improvement (-2.6 to +12.5) in active treatment groups found in double-blind knee arthritis studies over the past 30 years. No past studies could be found that quantified subjective swelling complaints or knee buckling frequency to compare with the 63% and 85% reductions demonstrated in the current study. Only 2 other studies indicated potential stabilization of radiograph findings similar to the current study.

**Previous Prolotherapy Injection Trials on Knee Ligament Laxity**

Double-blind studies of injection prolotherapy with non-inflammatory solutions for knee osteoarthritis or knee ligament laxity have not been previously reported. However, stimulation of the inflammatory cascade produces growth factors, and temporary inflammation induction by sodium morrhuate has been shown in a double-blind study in rabbits to thicken and strengthen knee collateral ligaments. The only human study on knee-ligament strengthening by inflammatory induction (using a 1.25% phenol 12.5% dextrose and 12.5% glycerine solution) had few patients and was unblinded. However, despite the low patient numbers, highly significant improvement of laxity measurements by a Genucom knee arthrometer was noted.

**Potential Applications and Future Study Implications**

This study is 1 of 2 concurrent double-blind studies (along with a concomitant finger arthritis study) to demonstrate that 10% dextrose alone is capable of a beneficial effect upon introduction into OA joints and that a treatment frequency of every 2 months is effective. Potential applications include patients too large or too young for total knee replacement, any patient in a third world country without replacement availability, patients who are symptomatic despite prescribed exercises or physical therapy or NSAIDs, or patients who are intolerant of NSAIDs. This is the first study to demonstrate in double-blind fashion that simple 10% dextrose will correct ACL ligament laxity in an objectively-measurable fashion. Potential applications may include patients with laxity without rupture, post surgical repair to prevent the typical post-surgical gradual loosening, and large total joint patients with dislocation tendency. The ability to intervene in a simple way for ACL laxity to limit the known complications of secondary arthritis should be of much interest. The broadness of application of dextrose injection in ligament/tendon treatment will depend on the cost of alternative treatments such as growth-factor-impregnated implants, direct stem cell injection, or injection of ACL ligament cells transfected with viruses whose genome has been altered to produce growth factors or to block growth factor inhibitors. The safety and low cost of dextrose injection may make it suitable for study in prophylactic use for knee injection in athletes prone to ACL injuries or in those with injuries but intact ligament.

These study results with 10% dextrose use are intriguing in that clinical experience indicates that dextrose 25% is superior to 10% dextrose in the treatment of knee OA and ACL laxity. This author is currently investigating the ability of patients to tell the difference between 10% and 25% dextrose upon injection into the knee in preparation for direct study of 25% dextrose, to be certain that double-blind protocols would not be affected by the brief inflammatory effect of 25% dextrose. Future study protocols using dextrose for prolotherapy should consider different volumes of dextrose injection, as some have suggested that smaller volumes are equally effective and may allow 25% dextrose to be used without patient awareness. Other applications of injection prolotherapy and areas of past and current study are covered in 2 recent publications.

If growth factor production results in more inexpensive and safe solutions for injection, this may be an alternative to stimulating growth factors by either brief inflammation or by dextrose
or by osmotic effects, and yet a likely outcome is that oral supplements, growth factor stimulant injection (prolotherapy), and direct growth factor provision by injection or other method will be complementary.

Frequency of treatment necessary for dextrose injection needs further evaluation, with current studies not designed to answer all questions about this. Clinical experience with 25% dextrose suggests that 2 to 3 bimonthly treatments are necessary prior to treatment taper.

For future studies on the ACL ligament, MRI availability to rule out complete ACL rupture and arthroscopy to confirm changes in cartilaginous surfaces would be ideal.

Now that the safety of dextrose in bacteriostatic water has been demonstrated in this study and a concomitant finger osteoarthritis study, future studies with dextrose should perhaps have dextrose in sterile water or saline versus an isotonic saline placebo.

Long-term radiographic follow-up data from the current study patients will be helpful to note net effect on cartilage and osteophytic change over a prolonged period, and patients are being followed for long-term radiographic findings.

CONCLUSIONS

Dextrose injection is clinically and statistically superior to bacteriostatic water in treatment of OA of the knee, with substantial improvements in joint pain, subjective joint swelling, flexion range of motion, and tendency for knee buckling. Anterior cruciate ligament tightening by objective measures was demonstrated with use of interarticular dextrose. Preliminary (1-year) radiographic findings show positive effects but 30- to 36-month followup radiography is planned for a clearer idea of the effect of proliferant injection on radiographic findings of OA. The inclusion of 38 knees in this study that were completely void of cartilage in at least 1 compartment, the long history of pain (8 years) in these knee OA patients, and their substantial improvements in joint pain, subjective joint swelling, prior to treatment taper.

Ger) OA, indicates that dextrose injection may have broad efficacies high glucose levels as a stimulus for expression of connective tissue growth factor and other genes in human mesangial cells. J Biol Chem. 1999;274(9):5830-5839.


Reinhold D, Ansong S, Schleicher ED. Elevated glucose levels stimulate transforming growth factor-beta 1 (TGF-beta 1) suppress interleukin IL-2, IL-6 and IL-10 production and DNA synthesis in peripheral blood mononuclear cells. Horm Metab Res. 1998;30(8):267-270.


